

AccuDiag™
T4
ELISA

Cat# 3149-16



Test	T4 ELISA
Method	Enzyme Linked Immunosorbent Assay
Principle	Competitive ELISA
Detection Range	0-30 µg/mL
Sample	50µL
Specificity	96.30%
Sensitivity	0.05 µg/mL
Total Time	~ 90 min
Shelf Life	12-14 Months from the manufacturing date

INTENDED USE

For the quantitative determination of thyroxine (T4) in human serum.

SUMMARY AND EXPLANATION

L-Thyroxine (T4) is a hormone that is synthesized and stored in the thyroid gland. Proteolytic cleavage of follicular thyroglobulin releases T4 into the bloodstream. Greater than 99% of T4 is reversibly bound to three plasma proteins in blood - thyroxine binding globulin (TBG) binds 70%, thyroxine binding pre-albumin (TBPA) binds 20%, and albumin binds 10%. Approximately 0.03% of T4 is in the free, unbound state in blood at any one time.

Diseases affecting thyroid function may present a wide array of confusing symptoms. Measurement of total T4 by immunoassay is the most reliable and convenient screening test available to determine the presence of thyroid disorders in patients. Increased levels of T4 have been found in hyperthyroidism due to Grave's disease and Plummer's disease and in acute and subacute thyroiditis. Low levels of T4 have been associated with congenital hypothyroidism, myxedema, chronic thyroiditis (Hashimoto's disease), and with some genetic abnormalities.

TEST PRINCIPLE

In the T4 EIA, a certain amount of anti-T4 antibody is coated on microtiter wells. A measured amount of patient serum, and a constant amount of T4 conjugated with horseradish peroxidase are added to the microtiter wells. During incubation, T4 and conjugated T4 compete for the limited binding sites on the anti-T4 antibody. After a 60 minutes incubation at room temperature, the wells are washed 5 times by water to remove unbound T4 conjugate. A solution of TMB is then added and incubated for 20 minutes, resulting in the development of blue color. The color development is stopped with the addition

of 2 N HCl, and the absorbance is measured spectrophotometrically at 450 nm. The intensity of the color formed is proportional to the amount of enzyme present and is inversely related to the amount of unlabeled T4 in the sample. By reference to a series of T4 standards assayed in the same way, the concentration of T4 in the unknown sample is quantified.

SPECIMEN COLLECTION AND PREPARATION

Serum should be prepared from a whole blood specimen obtained by acceptable medical techniques. This kit is for use with serum samples without additives only.

MATERIALS AND COMPONENTS

Materials provided with the test kits

1. Antibody-coated microtiter wells. 96 wells per bag.
2. Reference standard set, ready to use.
3. T4 HRPO Conjugate Diluent, 15 ml.
4. T4 HRPO Conjugate Concentrate, 0.8 ml
5. TMB Substrate, 12 ml.
6. Stop Solution, 12 ml.
7. Wash Buffer Concentrate (50X), 15ml

Materials required but not provided

1. Precision pipettes: 40µl~200µl and 1.0ml
2. Disposable pipette tips.
3. Distilled water.
4. Vortex mixer or equivalent.
5. Absorbent paper or paper towel.
6. Graph paper.
7. Microtiter well reader.

REAGENT PREPARATION

1. All reagents should be brought to room temperature (18-22°C) before use.
2. To prepare T4-HRPO Conjugate Reagent, add 0.1 ml of T4-HRPO Conjugate Concentrate to 2.0 ml of T4 Conjugate Diluent (1:20 dilution), and mix well. The amount of conjugate diluted is depend on your assay size. The Conjugate Reagent is stable at 4°C at least for two weeks.
Note: The T4 assay is a temperature sensitive assay. The best temperature condition for this assay is from 19°C to 22°C. If, in the environmental assay condition, the temperature is higher than expected, we recommend increasing the T4 conjugate dilution up to 1:40
3. Dilute 1 volume of Wash Buffer (50x) with 49 volumes of distilled water. For example, Dilute 15 ml of Wash Buffer (50x) into distilled water to prepare 750 ml of washing buffer (1x). Mix well before use.

ASSAY PROCEDURE

1. Secure the desired number of coated wells in the holder. Make data sheet with sample identification.
2. Dispense 50µl of standard, specimens, and controls into appropriate wells.
3. Dispense 100µl of Enzyme Conjugate Reagent into each well.
4. Thoroughly mix for 10 seconds. It is very important to have complete mixing in this step.
5. Incubate at room temperature (18-22°C) for 60 minutes.
6. Remove the incubation mixture by flicking plate contents into a waste container.
7. Rinse and flick the microtiter wells 5 times with washing buffer (1X).
8. Strike the wells sharply onto absorbent paper or paper towels to remove all residual water droplets.
9. Dispense 100µl of TMB solution into each well. Gently mix for 5 seconds.

10. Incubate at room temperature for 20 minutes without shaking.
11. Stop the reaction by adding 100µl of Stop Solution to each well. Gently mix for 5 seconds
12. Read optical density at 450nm with a microtiter well reader.

Important Note:

The wash procedure is critical. Insufficient washing will result in poor precision and falsely elevated absorbance readings.

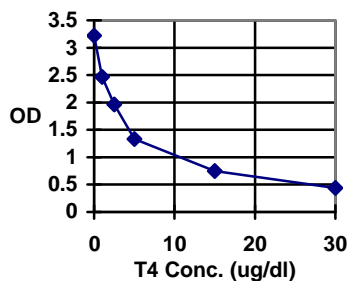
RESULTS

1. Calculate the average absorbance values (A₄₅₀) for each set of reference standards, control, and samples.
2. We recommend to use a proper software to calculate the results. If the software is not available, construct a standard curve by plotting the mean absorbance obtained for each reference standard against its concentration in µg/dl on linear graph paper, with absorbance on the vertical (y) axis and concentration on the horizontal (x) axis.
3. Using the mean absorbance value for each sample, determine the corresponding concentration of T4 in µg/dl from the standard curve.

Example of Standard Curve

Results of typical standard run with optical density reading at 450nm shown in the Y axis against T4 concentrations shown in the X axis. This standard curve is for the purpose of illustration only, and should not be used to calculate unknowns. Each user should obtain his or her own data and standard curve.

T4 (µg/dl)	Absorbance (450nm)
0	3.217
1	2.465
2.5	1.961
5	1.331
15	0.746
30	0.436



Expected Values and Sensitivity

T4 EIA was performed in a study of 200 euthyroid patients in one geographic location and yielded a **normal range of 5.0 to 13.0 µg/dl**. This range corresponds to those suggested by other commercial manufacturers. It is recommended that laboratories adjust values to reflect geographic and population differences specific to the patients they serve. The minimum detectable concentration of thyroxine by this assay is estimated to be 0.4µg/dl.

PERFORMANCE CHARACTERISTICS

I. Accuracy: Comparison between our kits and Commercial Available Kits provide the following data

N = 189
 Correlation Coefficient = 0.899
 Slope = 0.915
 Intercept = 1.635
 Mean (Our Assay) = 9.00
 Mean (Abbott) = 9.88

II Precision

1]. Intra-Assay:

Concentrations	Replicates	Mean	S.D.	% CV
Level I	20	1.92	0.199	10.36
Level II	20	6.78	0.403	5.95
Level III	20	19.46	1.09	5.59

2]. Inter-Assay:

Concentrations	Replicates	Mean	S.D.	% CV
Level I	20	2.05	0.285	13.9
Level II	20	8.51	0.594	7.0
Level III	20	19.78	1.16	5.8

III. Linearity

Two patient sera were serially diluted with 0 µg/dl standard in a linearity study. The average recovery was 106.9 %.

Sample A			
Dilution	Expected	Observed	% Recov.
undiluted	17.70	17.70	
2x	8.85	8.84	99.9
4x	4.43	4.42	99.8
8x	2.22	2.64	118.9
16x	1.11	1.37	123.4
<i>Average Recovery: 110.5 %</i>			

Sample B			
Dilution	Expected	Observed	% Recov.
undiluted	17.00	17.00	
2x	8.50	8.68	102.1
4x	4.25	4.51	106.1
8x	2.13	2.24	105.2
16x	1.07	1.07	100.0
<i>Average Recovery: 103.4 %</i>			

IV. Recovery

Various patient serum samples of known T4 levels were mixed and assayed in duplicate. The average recovery was 107.3 %.

Expected Concentration	Observed Concentration	% Recovery
17.9	19.6	109.0
8.3	9.2	110.8
4.6	4.9	106.5
2.0	2.2	110.0
0.6	0.6	100.0
<i>Average Recovery: 107.3 %</i>		

V. Sensitivity:


The sensitivity is defined as the concentration of T4 that corresponds to the absorbance that is two standard deviations greater than the mean absorbance of 20 replicates of the zero calibrator. The minimum detectable concentration of this assay is estimated to be 0.5 µg/dl.


VI. Cross-reactivity:

The following substances were tested for crossreactivity of the assay:

Antigens	Concentration	Equivalent T4	% Cross-Reactivity
Triiodothyroacetic Acid	100 µg/ml	2.9 µg/ml	2.90
Monoiodothyrosine	5,000 µg/ml	0.0 µg/ml	0.00
Diiodotyrosine	5,000 µg/ml	0.0 µg/ml	0.00
Methimazole	2,000,000 µg/ml	0.0 µg/ml	0.00
5,5'-Diphenylhydantoin	250,000 µg/ml	0.0 µg/ml	0.00
Phenylbutazone	1,000,000 µg/ml	0.0 µg/ml	0.00
6-n-propyl-2-thiouracil	25,000 µg/ml	0.0 µg/ml	0.00
Salicylic Acid	100,000 µg/ml	0.0 µg/ml	0.00
Acetylsalicylic Acid	5,000 µg/ml	0.3 µg/ml	0.006

**ISO 13485
ISO 9001**



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Date Adopted	Cat # 3149-16
2008-05-01	AccuDiag™- T4 ELISA-2013
<input type="checkbox"/> EC <input type="checkbox"/> REP	CEpartner4U, Esdoornlaan 13, 3951DB Maarn. The Netherlands. www.cepartner4u.eu
Revision Date: 10-01-2013	

PRECAUTIONS

STORAGE

- Unopened test kits should be stored at 2-8°C upon receipt and the microtiter plate should be kept in a sealed bag with desiccants to minimize exposure to damp air. The test kit may be used throughout the expiration date of the kit (One year from the date of manufacture). Refer to the package label for the expiration date.
- Opened test kits will remain stable until the expiring date shown, provided it is stored as prescribed above.
- A microtiter plate reader with a bandwidth of 10nm or less and an optical density range of 0-2 OD or greater at 450nm wavelength is acceptable for use in absorbance measurement.

REFERENCES

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