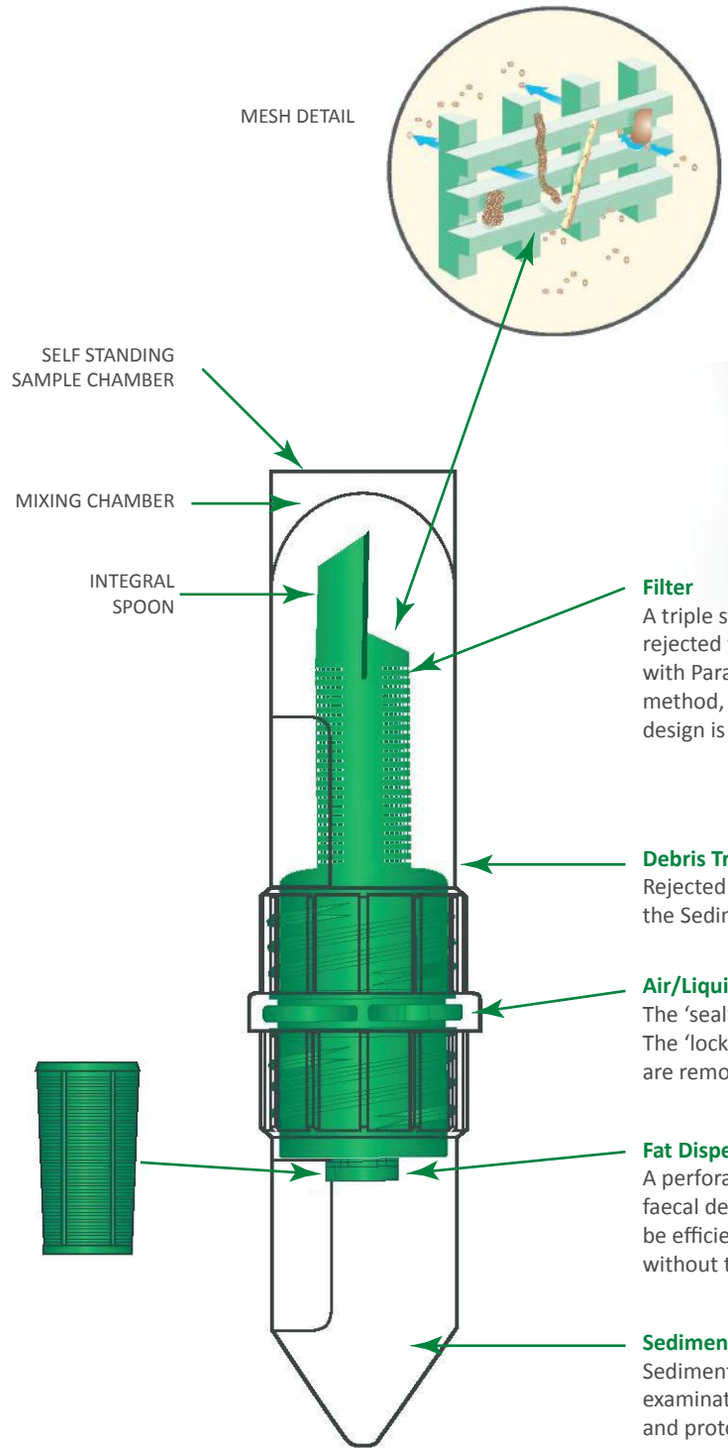
FOR FAECAL CONCENTRATION OF

HELMINTH OVA AND LARVAE / PROTOZOA CYSTS AND OOCYSTS



Filter
A triple stage matrix filtration. Large particles are rejected without obscuring filtration. Recovery rate with Parasep® is comparable to traditional sieve method, ie: Ridley-Allen. The vertical filter enclosed design is patented.

Debris Trap
Rejected particles are trapped to prevent extrusion into the Sedimentation Cone during centrifugation.

Air/Liquid Seal and Safety Lock
The 'seal' prevents the release of biohazardous material. The 'lock' ensures the Mixing Chamber and Filter are removed together for safe disposal.

Fat Dispersion Chamber
A perforated fat dispersion chamber removes the smaller faecal debris and separates the fat content so that it can be efficiently removed from the resulting sediment without the use of ether or ethyl acetate.

Sedimentation Cone
Sediment forms in the base of the cone allowing examination for the presence of helminth eggs or larvae and protozoa cysts or oocysts.

Health and Safety Benefits

- Totally enclosed/sealed process
- Reduced reagent volumes
- No cleaning required
- Single use, no sample contamination
- Ready to use systems available

Performance Benefits

- Optimum sample recovery
- Enhanced sample clarity
- Rapid four step process
- Human resources optimised
- Easy patient identification
- Fits all 50ml centrifuge buckets

PARASITOLOGY

SINGLE USE IN VITRO DIAGNOSTIC DEVICE

APACOR

Midi Parasep® SF

FAECAL PARASITE CONCENTRATOR

Midi Parasep® SF

FAECAL PARASITE CONCENTRATOR

Procedure

STEP 1 - SAMPLE PREPARATION

Fresh Samples

For empty Parasep®, unscrew lid and add 8.0ml of fixative and one drop of surfactant (eg: Apacor Triton X solution) to the mixing chamber.

Alternatively use the reagent ready Midi Parasep® SF.

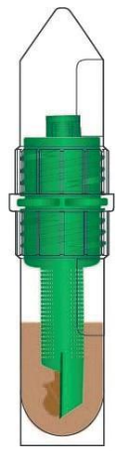
Introduce a level scoop of faecal sample to the fixative.

Mix in thoroughly with the Midi Parasep® SF spoon. If the sample is hard, break it up with the end of the spoon.

STEP 2 - EMULSIFICATION

Seal the Midi Parasep® SF by screwing in the filter/ sedimentation cone unit.

Vortex or shake to emulsify with the sedimentation cone pointing upwards.



STEP 3 - CENTRIFUGATION

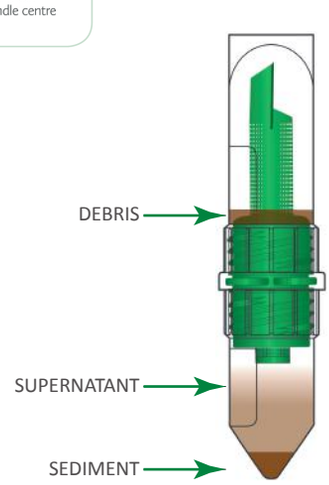
Invert the Midi Parasep® SF and centrifuge at 400g for two minutes
(J. Clin. Microbiol. doi:10.1128/JCM.00838-15).

Midi Parasep® SF fits all 50ml centrifuge buckets.

NOTE: TO CALCULATE THE REQUIRED RPM FOR ANY CENTRIFUGE.

$$RPM = \sqrt{\frac{g}{1.12r}} \times 1000$$

RPM - rotor speed in revs/min.
g - centrifugal force (max.1000g)
r - radius, horizontal distance between sedimentation cone tip and spindle centre measured in mm.

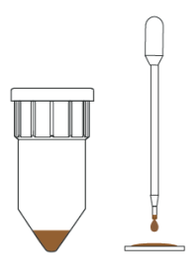


STEP 4 - EXAMINATION

Direct Method

Unscrew and discard the filter and mixing tube. Pour off all the liquid above the sediment.

Pipette one drop of sediment onto a slide, and cover with cover-slip. Alternatively, follow laboratory SOP for slide preparation.

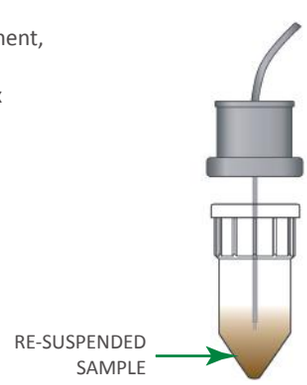


OR

Semi-automated System - ParaSys™

Unscrew and discard the filter and mixing tube. Pour off all the liquid above the sediment.

Insert aspirator into the sediment, press DILUTE to add saline to the sediment. Shake or vortex to re-suspend sample. Press SAMPLE to draw 100µl into the ParaSlide™.



See label for storage conditions and expiry date. Please adhere to the following guidelines when handling Midi Parasep® SF. To avoid cross contamination the Midi Parasep® SF device should remain closed at all times except when introducing the sample or when retrieving the final concentrated sample for examination.

Midi Parasep® SF is available empty or reagent ready
Please ask for details

Products can be ordered direct from Apacor or from an appointed distributor
Visit our website for all the latest information www.apacor.com or email on: sales@apacor.com

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